

FAT SCORING IN FOUR SPARROW SPECIES AS AN ESTIMATION OF BODY CONDITION: A VALIDATION STUDY

Elizabeth S. Wenker^{1}, Erin Kendrick, MS², Mike Maslanka, MS², and Michael L. Power, PhD¹*

¹*Center for Species Survival, Smithsonian National Zoological Park and Conservation Biology Institute, 2001 Connecticut Ave., NW Washington, DC 20008, USA.*

²*Department of Nutrition Sciences, Smithsonian National Zoological Park and Conservation Biology Institute, National Zoological Park, 3001 Connecticut Ave NW, Washington, DC 20008, USA.*

Abstract

Body condition indices are used to assess individual health of wild and captive animals. The definition of body condition varies by researcher and study goals but typically refers to measures of energy reserves; most commonly fat stores (Labocha & Hayes, 2012). In avian biology, body condition has been correlated with individual survivability (Blums *et al.*, 2005), reproduction (Chastel *et al.*, 1995; Bêty *et al.*, 2003) migration (Bêty *et al.*, 2003; McWilliams *et al.*, 2004; Laursen *et al.*, 2019), and habitat quality (Angelier *et al.*, 2011; Balbontín *et al.*, 2012). One of the oldest and most common methods of determining avian body condition is fat scoring: using a qualitative scale to score visible subcutaneous fat (Blanchard, 1941; Helms & Drury, 1960). Scores are determined by using the fullness and color of furcular and/or abdominal regions of a bird to estimate fat reserve size and can be determined in under a minute. Fat pad size and fat score are highly correlated, making this a quick and effective means of determining body condition (Kaiser, 1993; Labocha & Hayes, 2012). Fat score can be used in tandem with other morphological measurements to more accurately predict fat mass (Labocha & Hayes, 2012; McWilliams & Whitman, 2013). However, it is important to note that fat score is a qualitative measurement, and therefore subjective, and there is not one single scale used (Rogers, 2003; Labocha & Hayes, 2012; McWilliams & Whitman, 2013). Furthermore, not all bird species carry fat in the same manner (Seewagen, 2008; Schamber *et al.*, 2009).

The only accurate way to determine the lipid composition of a bird (including non-fat pad lipids) is to chemically extract it from the body, which is fatal to the bird and time-consuming for researchers. The number of studies using such methodologies is limited due to the ethical nature of killing subjects but exist for a few species of passerines and waterfowl (Conway *et al.*, 1994; Seewagen, 2008; Schamber *et al.*, 2009; McWilliams & Whitman, 2013; Beuth *et al.*, 2016) The goal of this study is to determine the relationship between fat score and total body lipid composition via chemical extraction in four sparrow species: white-throated sparrows (*Zonotrichia albicollis*), song sparrows (*Melospiza melodia*), swamp sparrows (*Melospiza georgiana*), and Lincoln's sparrows (*Melospiza lincolnii*).

Methods

A total of 42 white-throated (hereafter referred to as WTSP), 19 song, 5 swamp, and 3 Lincoln's sparrows were collected opportunistically by City Wildlife as part of their Lights Out DC initiative in the spring and fall seasons of 2017-2019. All birds died due to building collisions and were donated to the Smithsonian National Zoological Park's (SI-NZP) Nutrition Department.

Visual Assessment

Birds were thawed and scored using the ESF system from the British Trust for Ornithology (0 - 8). The same procedures were followed as if scoring live birds and scoring was assumed to be the same as if done on live specimens (Krementz & Pendleton, 1990). To reduce inter-observer variation, all scoring was performed by one SI-NZP zoo nutritionist with extensive body scoring experience.

Fat Determination

Initial body weight was measured for all birds. Individuals were defeathered manually and dissected ventrally to remove their fat pads as completely as possible. Beaks were removed via scalpel, and legs were removed via severance at or just above the intertarsal joint to aid in the homogenizing process (note: lipid content in these areas are absent or negligible). Birds were then reweighed. The altered bodies were then blended with distilled water in a household blender until a homogenized slurry was produced. The slurries were dried in aluminum pans at 100°C in forced air convection oven for 24 hours and then manually homogenized into a powder. Crude fat (CF) content of the fat pads (abdominal and furcular combined per bird) and the homogenized carcass powder was measured using an ANKOM fat apparatus (ANKOM XT15 Extractor, Macedon, NY).

Results

All birds measured within the standard size and/or weight range for their species (Ammon, 2020; Arcese *et al.*, 2020; Falls & Kopachena, 2020; Herbert & Mowbray, 2020) All birds scored within the 0.5-4 range, which falls in the lower half of the fat score scale and is indicative of a typical wild population (Witter & Cuthill, 1993). There was a wide range in total lipid content across the birds. The average % total lipid content in order of greatest to least was seen in swamp sparrows, song sparrows, WTSP, and then Lincoln's sparrows (32.8 ± 5.3 , 29.2 ± 2.1 , 27.6 ± 1.4 , and $22.6\pm 7.3\%$, respectively), but the ranges and sample sizes varied greatly. For the fat pads themselves, the average %CF ranged from 79.7-96.4% across species, with individual values ranging from 39% to 100%.

Fat score was not related to body mass in WTSP but was related to all of the percent fat values. Body mass was not correlated with carcass fat or total fat and, interestingly, negatively correlated with fat pad percent fat. The correlations for song sparrows were similar except that in this species body mass was correlated with measures of body fat. Correlations were not done on the swamp and Lincoln's sparrows due to small sample size, but their values follow the pattern for WTSP, with fat score appearing to be associated with measures of fat but not necessarily with body mass.

Discussion

Overall, our results indicated that fat scoring is a valid method for predicting body condition of these four sparrow species. This is consistent with literature regarding other small passerines (Conway *et al.*, 1994; Stevenson & Woods, 2006; Seewagen, 2008; Labocha & Hayes, 2012).

Low percent fat in fat pads was found in birds with low body condition scores (0.5 or 1). Fat pads contain non-fat material (vascular and connective tissue) and dissection may remove some surrounding tissue, which possibly represented a greater proportion of tissue with small fat pads compared to large pads which routinely had high percentage fat values.

The average lipid content of the carcass ranged from 20.9-31.6% across species. The lowest percentage was 6.2% in a song sparrow (fat score = 0.5) and the highest 48.8% in a WTSP (fat score = 4). Upon dissection, additional lipid stores were primarily seen lining the intestines, at the shoulder joint, and above the caudal vertebrae. It is important for researchers to understand that while the subcutaneous fat pads are the primary lipid reservoirs of birds, mesenteric fat can play an important part when these stores are depleted.

Conclusion

Fat scoring offers a quick method of accurately assessing body condition in small passerines like these sparrows but does not provide exact lipid content.

Literature cited

- Ammon EM (2020) Lincoln's Sparrow (*Melospiza lincolnii*). In Poole AF and Gill FB, Eds. Birds of the World.
- Angelier F, Tonra CM, Holberton RL, and Marra PP (2011) Short-term changes in body condition in relation to habitat and rainfall abundance in American redstarts *Setophaga ruticilla* during the non-breeding season. *J Avian Biol* 42(4): 335–341.
- Arcese P, Sogge MK, Marr AB, and Patten MA (2020) Song Sparrow (*Melospiza melodia*). In Poole AF and Gill FB, Eds. Birds of the World.
- Balbontín J, Pape Møller A, Hermosell IG, Marzal A, Reviriego M, and De Lope F (2012) Lifetime individual plasticity in body condition of a migratory bird. *Biol J Linn Soc Lond* 105(2): 420–434.
- Bêty J, Gauthier G, and Giroux JF (2003) Body condition, migration, and timing of reproduction in snow geese: A test of the condition-dependent model of optimal clutch size. *Am Nat* 162(1): 110–121.
- Beuth JM, Paton PWC, Osenkowski JE, and McWilliams SR (2016) Validating the deuterium dilution method to measure body composition of common eider. *Wildl Soc Bull* 40(3): 456–463.
- Blanchard BD (1941) The white-crowned sparrows (*Zonotrichia leucophrys*) of the Pacific seaboard: environment and annual cycle. *Univ Calif Publ Zool* 46: 1-178.
- Blums P, Nichols J, Hines J, Lindberg M, and Mednis A (2005) Individual quality, survival variation and patterns of phenotypic selection on body condition and timing of nesting in birds. *Oecologia* 143(3): 365–376.
- Chastel O, Weimerskirch H, and Jouventin P (1995) Influence of body condition on reproductive decision and reproductive success in the blue petrel. *Auk* 112(4): 964–972.
- Conway CJ, Eddleman WR, and Simpson KL (1994) Evaluation of lipid indices of the wood thrush. *Condor* 96(3): 783–790.

- Falls JB and Kopachena JG (2020) White-throated Sparrow (*Zonotrichia albicollis*). In: Poole AF, Ed. Birds of the World.
- Helms CW and Drury WH (1960) Winter and migratory weight and fat field studies on some North American buntings. *Bird-Banding* 31(1): 1–40.
- Herbert JA and Mowbray TB (2020) Swamp Sparrow (*Melospiza georgiana*). In: Rodewald PG, Ed. Birds of the World.
- Kaiser A (1993) A new multi-category classification of subcutaneous fat deposits of songbirds. *J Field Ornithol* 64(2): 246–255.
- Labocha M and Hayes JP (2012) Morphometric indices of body condition in birds: a review. *J Ornithol* 153(1): 1–22.
- Laursen K, Møller AP, Haugaard L, Öst M, and Vainio J (2019) Allocation of body reserves during winter in eider *Somateria mollissima* as preparation for spring migration and reproduction. *J Sea Res* 144: 49–56.
- McWilliams SR, Guglielmo C, Pierce B, Klaassen M (2004) Flying, fasting, feeding in birds during migration: a nutritional physiology ecology perspective. *J Avian Biol* 35(5): 377–393.
- McWilliams S R and Whitman M (2013) Non-destructive techniques to assess body composition of birds: a review and validation study. *J Ornithol* 154(3): 597–618.
- Rogers CM (2003) New and continuing issues with using visible fat classes to estimate fat stores of birds. *J Avian Biol* 34(2): 129–133.
- Schamber JL, Esler D, and Flint PL (2009) Evaluating the validity of using unverified indices of body condition. *J Avian Biol* 40(1): 49–56.
- Seewagen CL (2008) An evaluation of condition indices and predictive models for noninvasive estimates of lipid mass of migrating Common Yellowthroats, Ovenbirds, and Swainson's Thrushes. *J Field Ornithol* 79(1): 80–86.
- Stevenson RD and Woods WA (2006) Condition indices for conservation: new uses for evolving tools. *Integr Comp Biol* 46(6): 1169–1190.
- Witter MS and Cuthill IC (1993) The ecological costs of avian fat storage. *Philos Trans R Soc Lond B Biol Sci* 340(1291): 73–92.